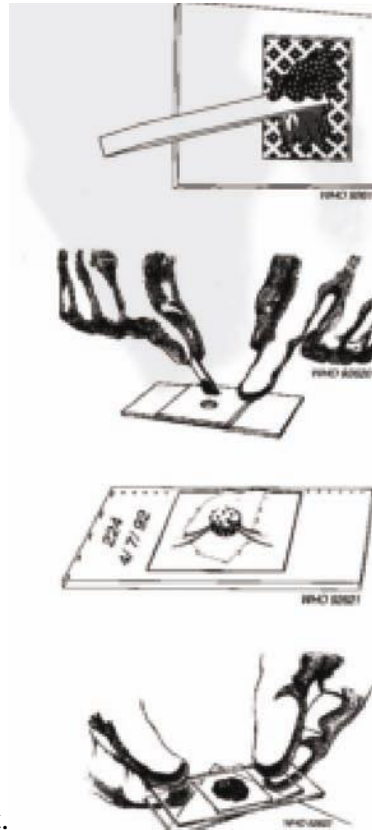


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| SOP Kato-Katz | |
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| Parameter: | Stool samples |
| Method: | Kato-Katz |
| Principle: | <p>Kato Katz technique is used for qualitative and semi-quantitative diagnosis of intestinal helminthic infestations; caused by <i>Ascaris lumbricoides</i>, <i>Trichuris trichiura</i>, hookworm and <i>Schistosoma spp.</i> WHO has recommended Kato Katz technique in areas with moderate to high transmission rates of soil transmitted helminths (i.e. where the proportion of infected individuals is >20– >50%) or intestinal schistosomiasis (>10–50%)</p> <p>People infected with STH or intestinal schistosomes pass the eggs of the worms through their faeces. In the Kato-Katz technique faeces are pressed through a mesh screen to remove large particles. A portion of sieved sample is then transferred to the hole of a template on a slide. After filling the hole, the template is removed and the remaining sample is covered with a peice of cellophane soaked in glycerol. The glycerol clears the faecal material from around the eggs. The eggs are then counted and the number calculated per gram of faeces.</p> <p>taken from https://microbeonline.com/kato-katz-technique-principle-procedure-results/</p> |
| Sample: | Proband stool sample |
| Sample storage prior to analysis: | Room temperature |
| Analyser: | microscope |
| Reagents (with source of supply) | <ul style="list-style-type: none"> • Kato-set (Template with hole, screen, nylon or plastic, plastic spatula) • Newspaper or glazed tile • Microscope slides • Cellophane as cover slip, soaked in Glycerol-malachite green or or glycerol-methylene blue solution. • Fresh stool • Gloves |
| Method implementation: | <ol style="list-style-type: none"> 1. Label a glass slide with the sample number and then place a plastic template on top of |



- it.
2. Place a small amount of the faecal sample on a newspaper and press a piece of nylon screen on top. Using a spatula, scrape the sieved faecal material through the screen so that only the debris remains.
 3. Scrape up some of the sieved faeces to fill the hole in the template, avoiding air bubbles and levelling the faeces off to remove any excess.
 4. Carefully lift off the template and place it in a bucket of water mixed with concentrated detergent so that it can be reused.
 5. Place one piece of the cellophane, which has been soaked overnight in methylene blue glycerol solution, over the faecal sample.
 6. Place a clean slide over the top and press it evenly downwards to spread the faeces in a circle. Carefully remove the slide by gently sliding it sideways to avoid separating the cellophane strip. If done well, it should be possible to read newspaper print through the stool smear. Place the slide with the cellophane upwards.

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| Criteria for the analysis of results: | Note: If hookworm is present in the area the slide should be read within 30–60 minutes. After that time, the hookworm eggs disappear. |
| Evaluation and documentation | Probands which tested positive for intestinal helminths should get a remark on their file |
| Information on method validation: | <p>WHO has recommended Kato Katz technique in areas with moderate to high transmission rates of soil transmitted helminths (i.e. where the proportion of infected individuals is >20– >50%) or intestinal schistosomiasis (>10–50%). Where the prevalence of soil transmitted helminths (STH) is <20%, the specificity of this technique makes it less appropriate, and more sensitive tools should be used.</p> <p>Taken from: https://microbeonline.com/kato-katz-technique-principle-procedure-results/</p> |
| Literature: | <p>Taken from: https://microbeonline.com/kato-katz-technique-principle-procedure-results/</p> |